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    ANSWER 1 OF 2 HCAPLUS COPYRIGHT 2009 ACS on STN DUPLICATE 1
    2005:300594 HCAPLUS
ΑN
    142:368184
DN
    Production of biol. active polypeptides by the proteolysis of recombinant
ΤТ
    synthetic polypeptide precursors by the OmpT protease variants
    Okuno, Kazuaki; Yabuta, Masayuki
IN
    Daiichi Suntory Pharma Co., Ltd., Japan
PA
SO
    PCT Int. Appl., 107 pp.
    CODEN: PIXXD2
DT
    Patent
    Japanese
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                       KIND DATE
    PATENT NO.
                                         APPLICATION NO.
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                        A1 20050407 WO 2004-JP14704
    WO 2005030956
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                                        EP 2004-773628
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PRAI JP 2003-342183
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    WO 2004-JP14704
                               20040929
ASSIGNMENT HISTORY FOR US PATENT AVAILABLE IN LSUS DISPLAY FORMAT
    The proteolytic method for producing biol. active polypeptides (
    ACTH (1-24), motilin or calcitonin) from recombinant
    synthetic precursor polypeptides or fusion proteins by using OmpT
    protease mutants has been developed. The synthetic precursor
    polypeptides or fusion proteins (22 .apprx. 45 a.a. (amino acid)) have
    been designed according to the substrate specificities of the OmpT
    protease mutants. Synthetic substrate polypeptides have Arg or
    Lys at P1 site and the a.a. other than Asp, Glu or Pro at the P1' site.
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The substrate polypeptides have one, two or serial three basic a.a. in the

P10 .apprx. P3, P10 .apprx. P3' or P10 .apprx. P5' (more specifically in the P5 .apprx. P3 site), however the sites P6 and P4 are excluded if only one basic a.a. in the sequence. The fusion protein substrates with protection peptide having C-terminal Arg or Lys have N-terminal a.a. such as Phe, Ala, Ser, Cys or Tyr and the other a.a. excluding Asp, Glu and Pro. These preferred P5 .apprx. P1 sequence and P7 .apprx. P1 sequence in the synthetic precursor polypeptides or fusion proteins are Arg-Arg-Arg-Ala-Arg and Asp-Ala-Arg-Arg-Arg-Ala-Arg, resp. Introduction of acidic a.a. typically Asp to the P3 site can repress the digestion by the OmpT proteases. The OmpT protease variants that can be used in the proteolysis system have a.a variation at the 97th position. The 97th a.a. is Leu, Met or His and the other a.a. including Ala, Phe, Ser, Thr, Cys, Asn, Gln, and Glu. The vector encoding the fusion substrate protein containing human glucagon, motilin, ACTH or calcitonin was designed to satisfy the structural condition claimed above and expressed in the inclusion body of E. coli and the cleaving of biol. active peptides from the substrate fusion proteins by the recombinant OmpT protease variant was demonstrated. The performance of the coexpression system of the substrate fusion protein and OmpT protease variant in the biol. active peptide generation was also demonstrated.

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L3 ANSWER 2 OF 2 MEDLINE on STN

DUPLICATE 2

AN 2004028684 MEDLINE

DN PubMed ID: 14711628

- TI Utilization of Escherichia coli outer-membrane endoprotease OmpT variants as processing enzymes for production of peptides from designer fusion proteins.
- AU Okuno Kazuaki; Yabuta Masayuki; Ooi Toshihiko; Kinoshita Shinichi
- CS Institute for Medicinal Research and Development, Daiichi Suntory Pharma Co., Ltd., Akaiwa, Chiyoda-machi, Ohra-gun, Gunma 370-0503, Japan.. Kazuaki\_Okuno@dsup.co.jp
- SO Applied and environmental microbiology, (2004 Jan) Vol. 70, No. 1, pp. 76-86.

Journal code: 7605801. ISSN: 0099-2240.

Report No.: NLM-PMC321264.

CY United States

- DT Journal; Article; (JOURNAL ARTICLE)
- LA English
- FS Priority Journals
- EM 200404
- ED Entered STN: 21 Jan 2004
  Last Updated on STN: 9 Apr 2004
  Entered Medline: 8 Apr 2004
- Escherichia coli outer-membrane endoprotease OmpT has suitable properties AΒ for processing fusion proteins to produce peptides and proteins. However, utilization of this protease for such production has been restricted due to its generally low cleavage efficiency at Arg (or Lys)-Xaa, where Xaa is a nonbasic N-terminal amino acid of a target polypeptide. The objective of this study was to generate a specific and efficient OmpT protease and to utilize it as a processing enzyme for producing various peptides and proteins by converting its substrate specificity. Since OmpT Asp(97) is proposed to interact with the P1' amino acid of its substrates, OmpT variants with variations at Asp(97) were constructed by replacing this amino acid with 19 natural amino acids to alter the cleavage specificity at Arg (P1)-Xaa (P1'). The variant OmpT that had a methionine at this position, but not the wild-type OmpT, efficiently cleaved a fusion protein containing the amino acid sequence -Arg-Arg-Arg-Ala-Arg downward arrow motilin, in which

motilin is a model peptide with a phenylalanine at the N terminus. The OmpT variants with leucine and histidine at position 97 were useful in releasing human adrenocorticotropic hormone (1-24) (serine at the N terminus) and human calcitonin precursor (cysteine at the N terminus), respectively, from fusion proteins. Motilin was produced by this method and was purified up to 99.0% by two chromatographic steps; the yield was 160 mg/liter of culture. Our novel method in which the OmpT variants are used could be employed for production of various peptides and proteins.